

# Validação de métodos em diagnóstico molecular

Gustavo Barra



Essa é mais uma iniciativa educativa da

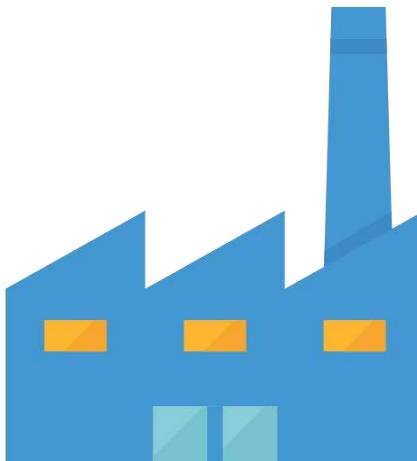
# Homenagem e agradecimento

Dr. Marcio Mendes Biasolli  
Família Controllab



Ilustração livro Paí, Paí vini-çá  
Autor: Marcio Mendes Biasolli

# Conceitos e definições

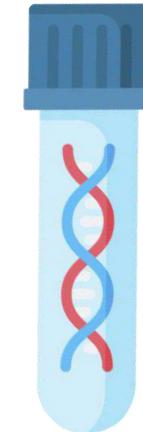


Verificação  
E, P, LOD, IR,



## Validação

- E, P, LOD, LOQ, IR
- Comparação
- Recuperação
- Clínica
- Em andamento
- Baseada no método



## Diagnóstico Molecular

- Detectar
- Tipar
- Quantificar
- DNA/RNA

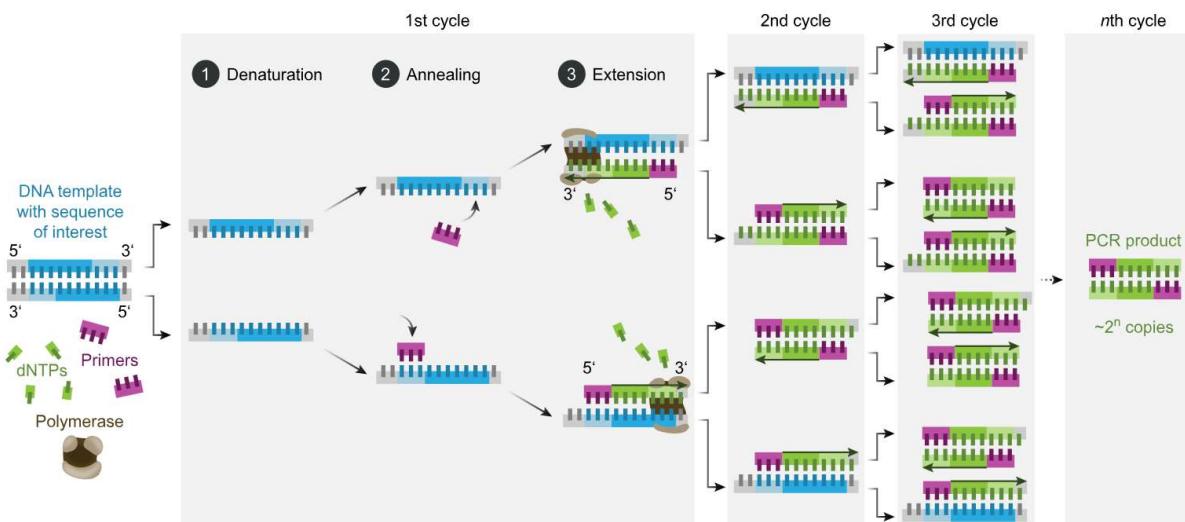


## Teste in-house (LDT)

- Novo/modificado
- Exclusivos
- Validade clínica
- PCR, NGS, HPLC, MALDI-TOF

# Diagnóstico molecular

## qPCR

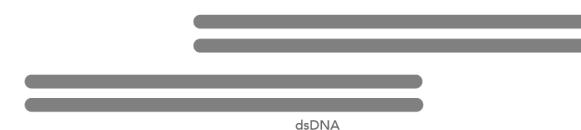


Cq

Sci 2022, 4(1), 4; <https://doi.org/10.3390/sci4010004>

## NGS

Fragmentation



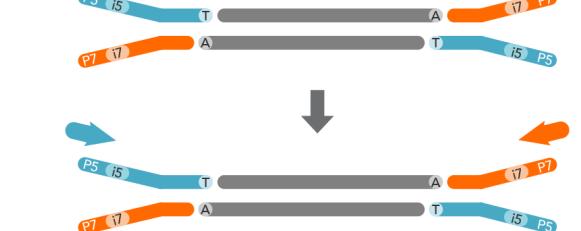
End repair and A-tailing



Ligation



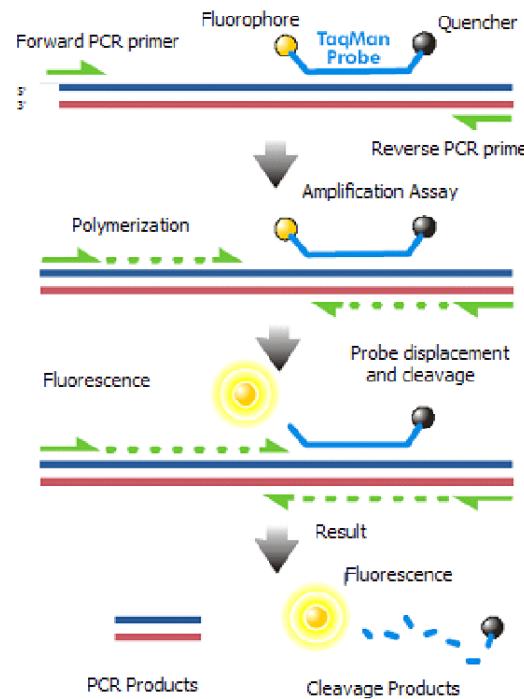
PCR amplification



ACTGCTGA

# Diagnóstico molecular

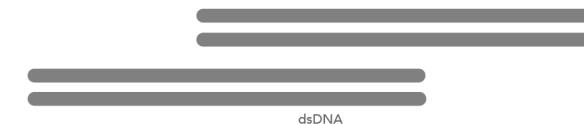
## qPCR



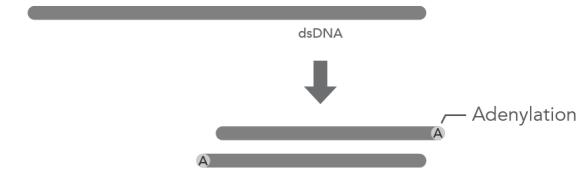
Cq

## NGS

### Fragmentation



### End repair and A-tailing



### Ligation

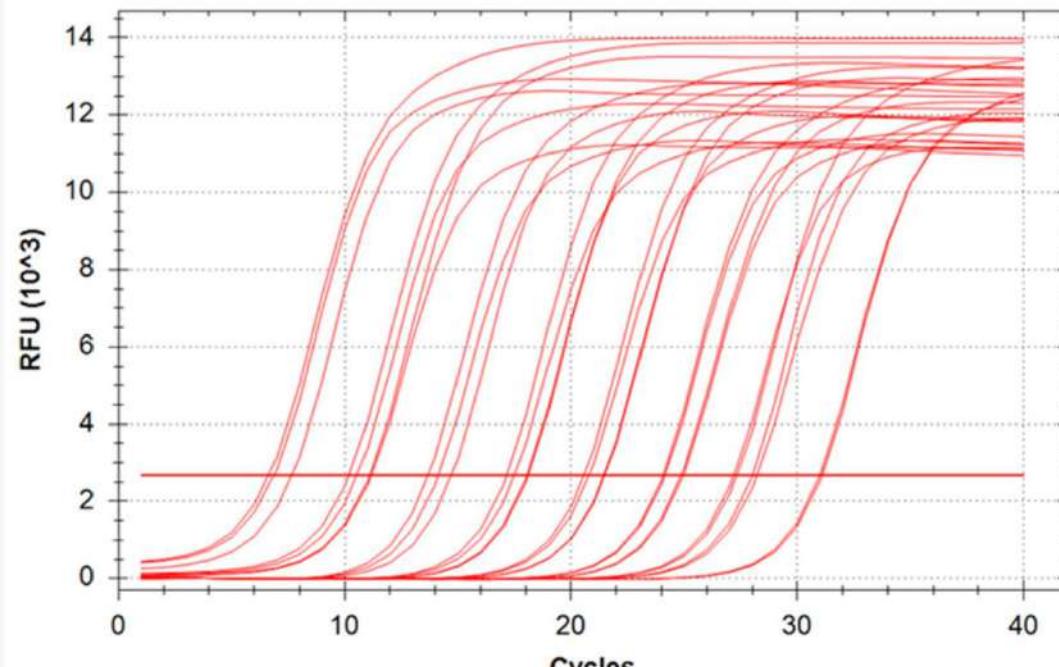


### PCR amplification

ACTGCTGA

# Diagnóstico molecular

qPCR



Cq

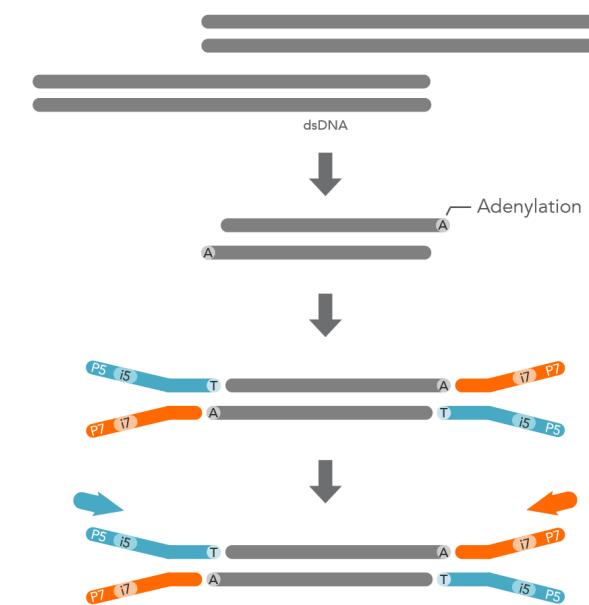
NGS

Fragmentation

End repair and A-tailing

Ligation

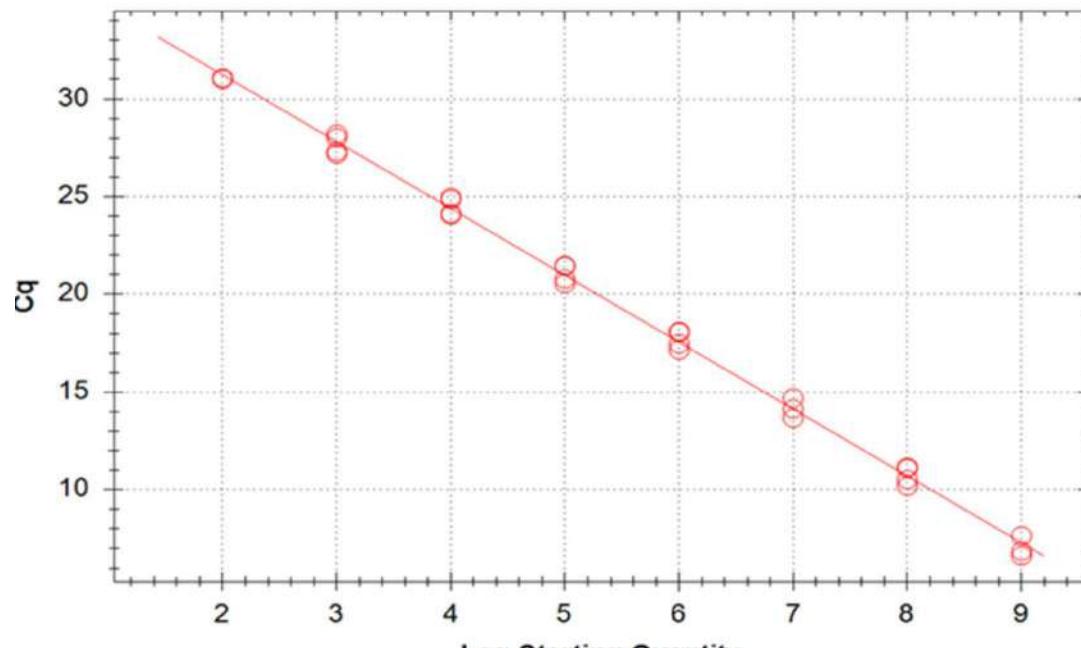
PCR amplification



ACTGCTGA

# Diagnóstico molecular

## qPCR



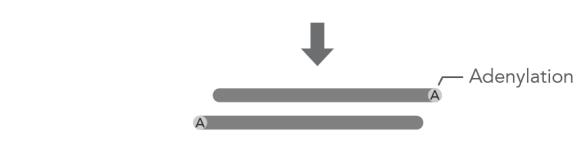
Cq

## NGS

Fragmentation



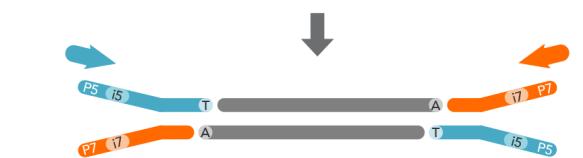
End repair and A-tailing



Ligation



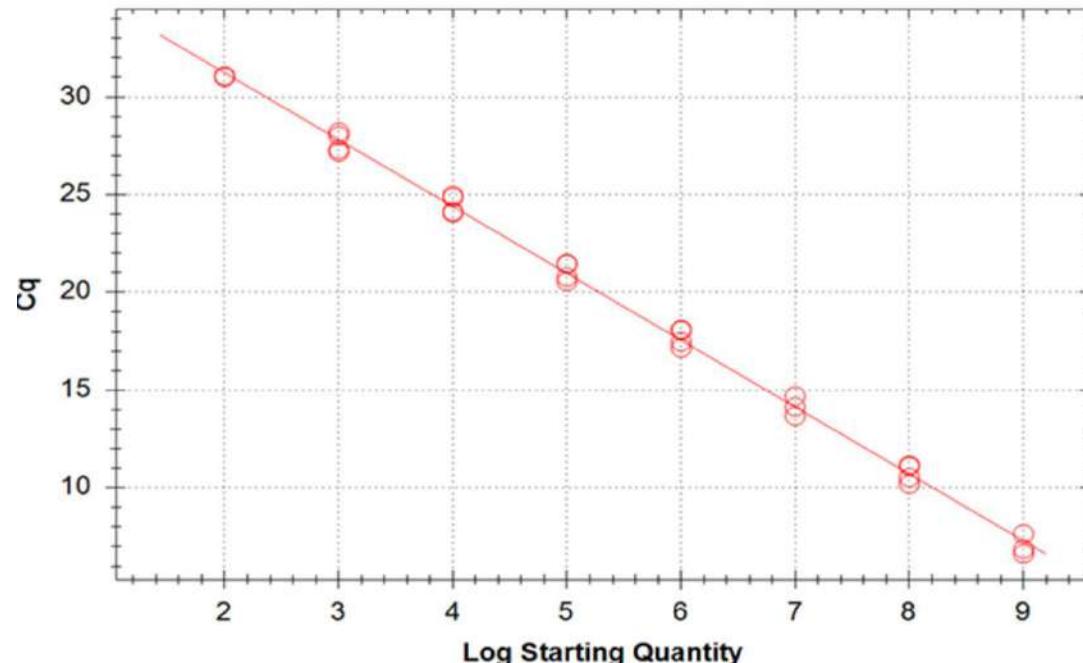
PCR amplification



ACTGCTGA

# Diagnóstico molecular

qPCR



Cq

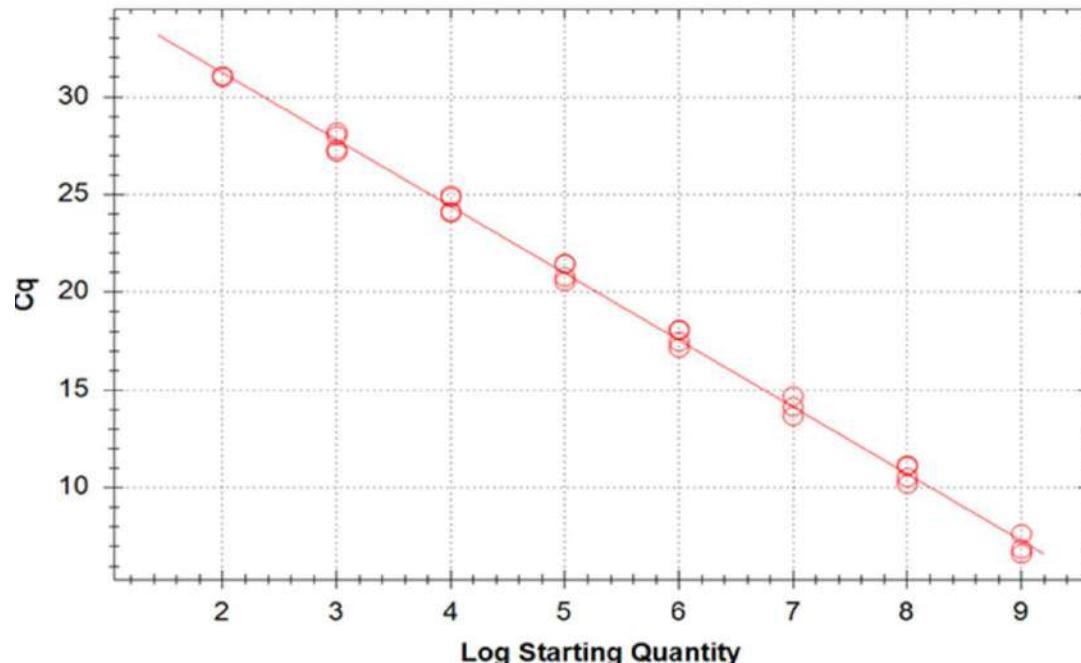
NGS



ACTGCTGA

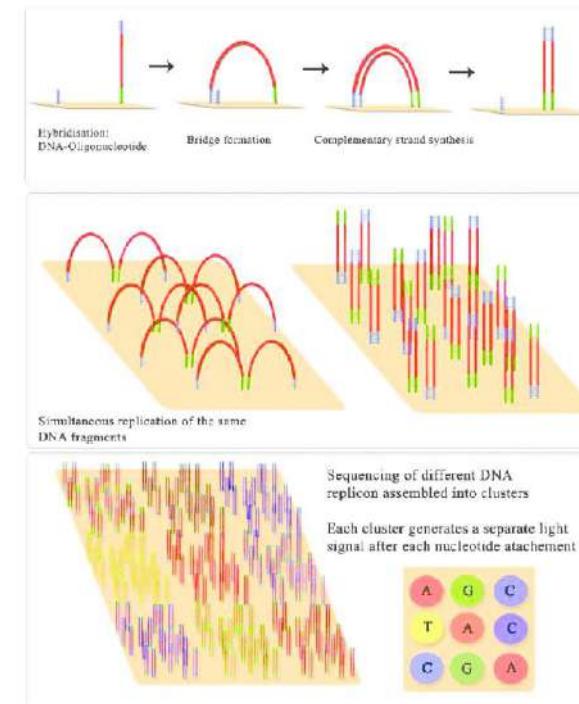
# Diagnóstico molecular

## qPCR



Cq

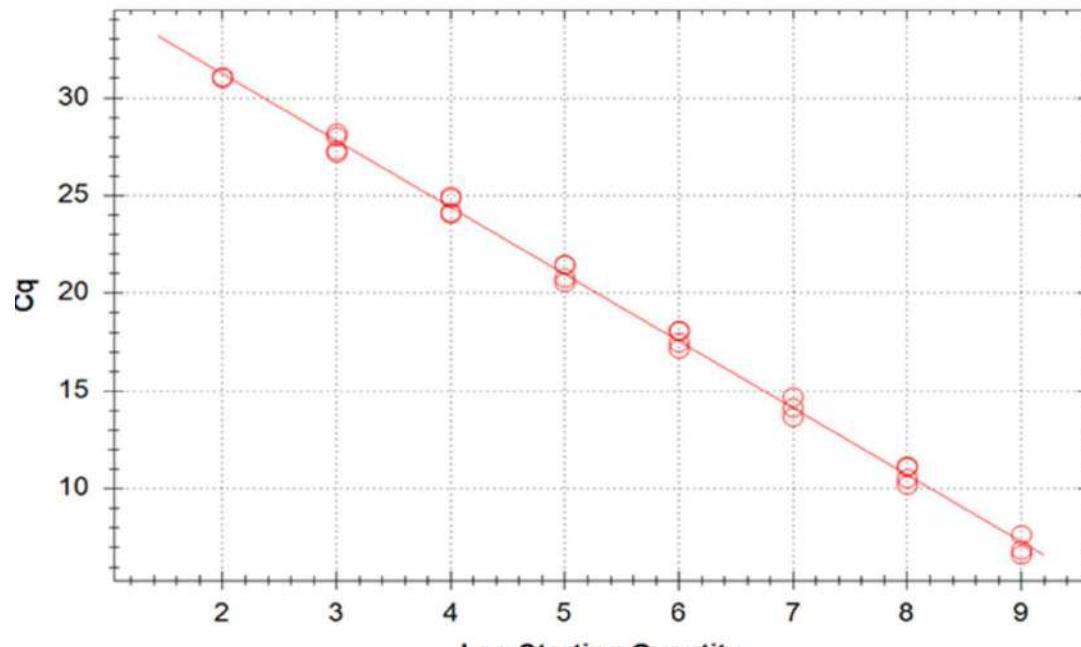
## NGS



ACTGCTGA

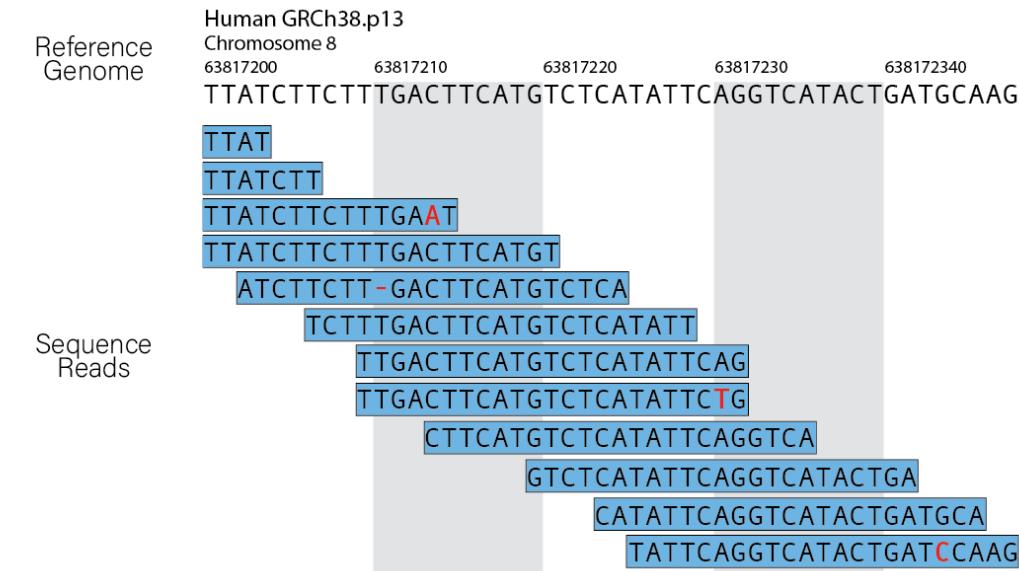
# Diagnóstico molecular

qPCR



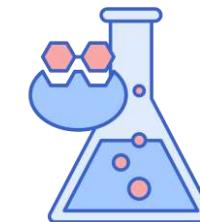
Cq

NGS

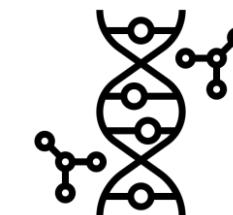


ACTGCTGA

# Diagnóstico molecular

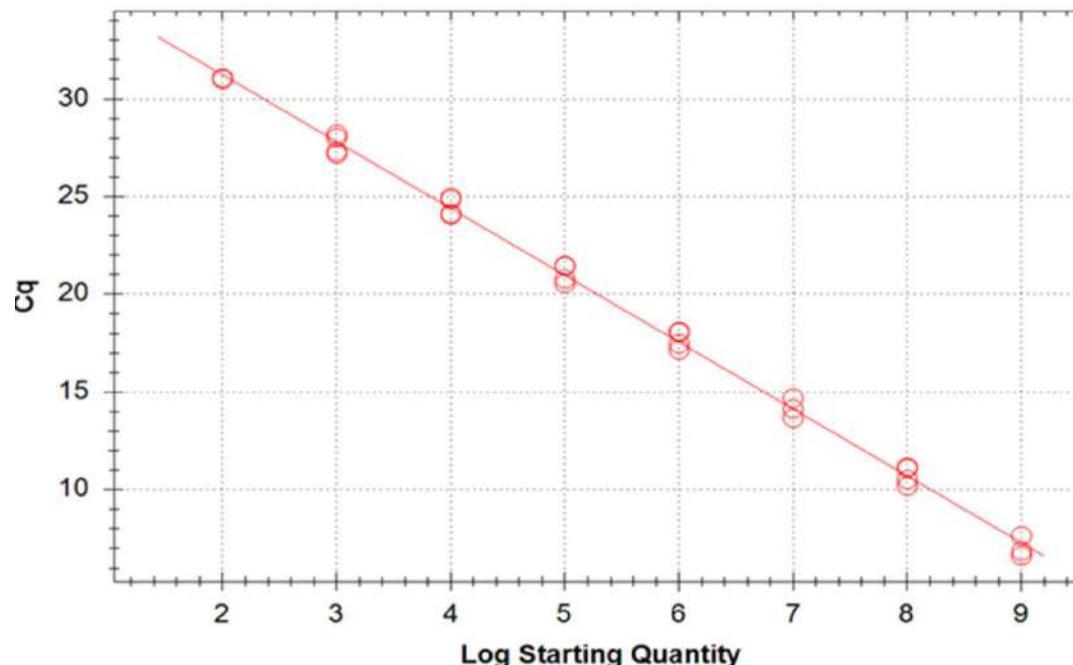


**Enzimologia**



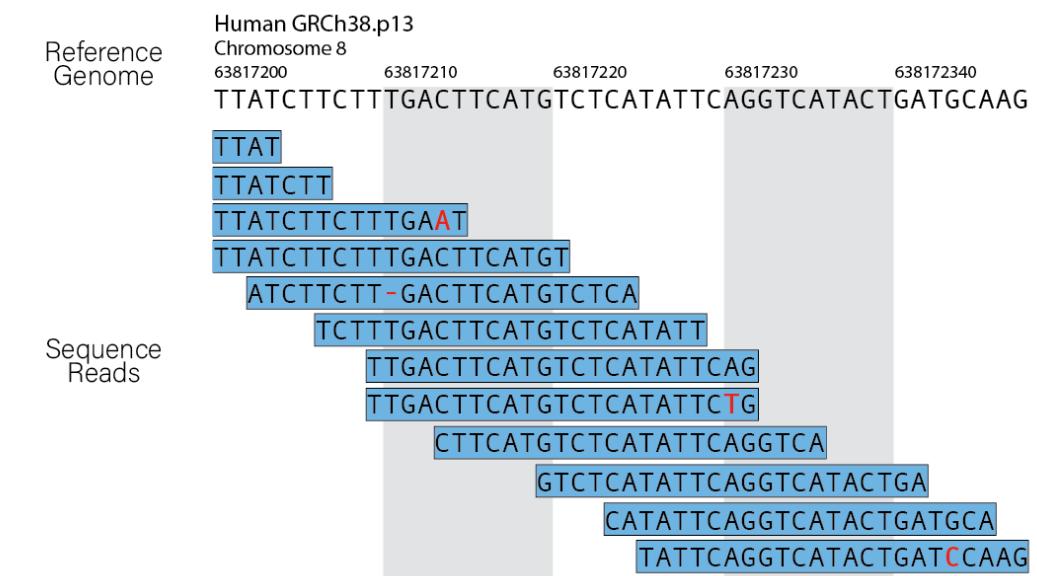
**Biología sintética**

**qPCR**



**Cq**

**NGS**



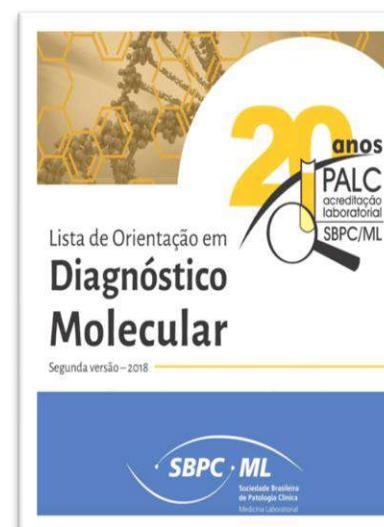
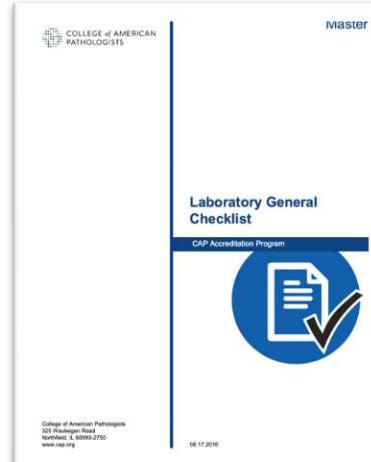
**ACTGCTGA**

# Regras do jogo – métodos próprios (LDT)

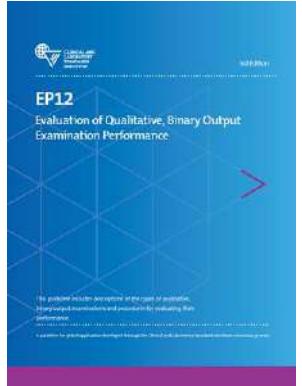
Regulamentação  
“Autoriza a fazer”



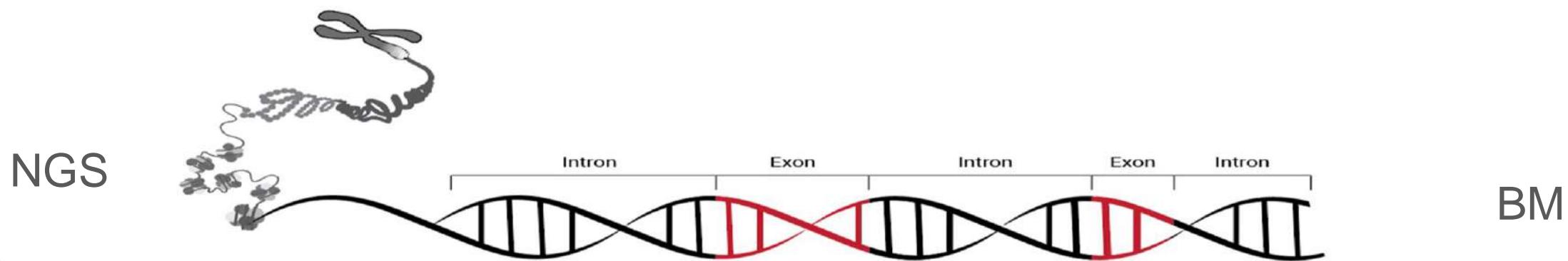
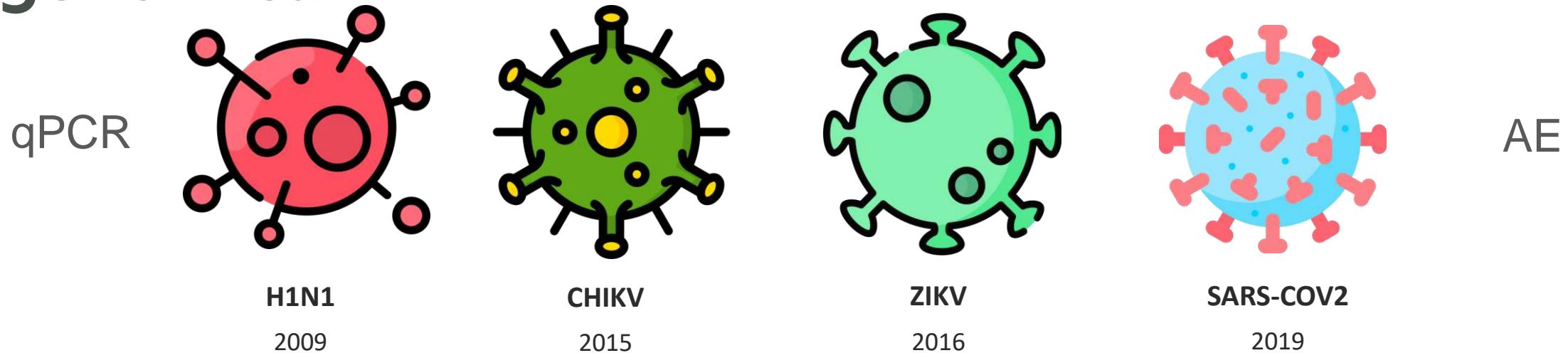
Certificações  
“O que fazer”



Guidelines e referências  
“Como fazer”

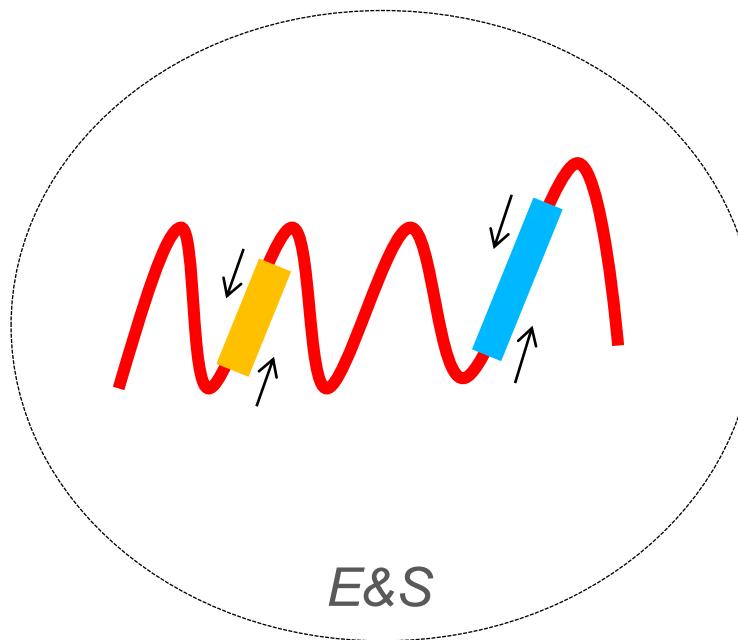


# Métodos próprios - Surtos, epidemias e genômica



# Ensaios independentes

qPCR

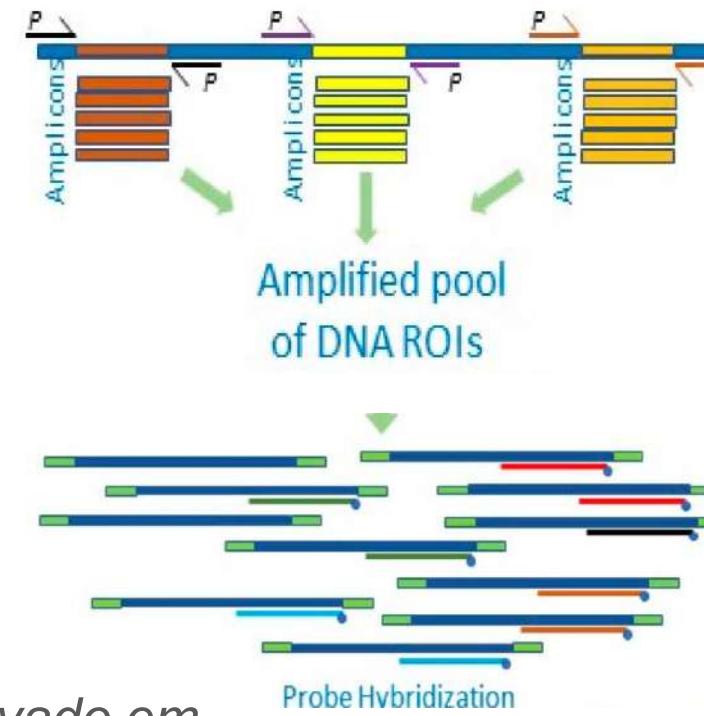


Primers

Concordância  
*T, P, e N*

*“Se mesmo resultado for observado em pelo menos dois ensaios independentes e o método destes ensaios já é estabelecido o resultado é considerado verdadeiro”*

NGS

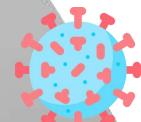
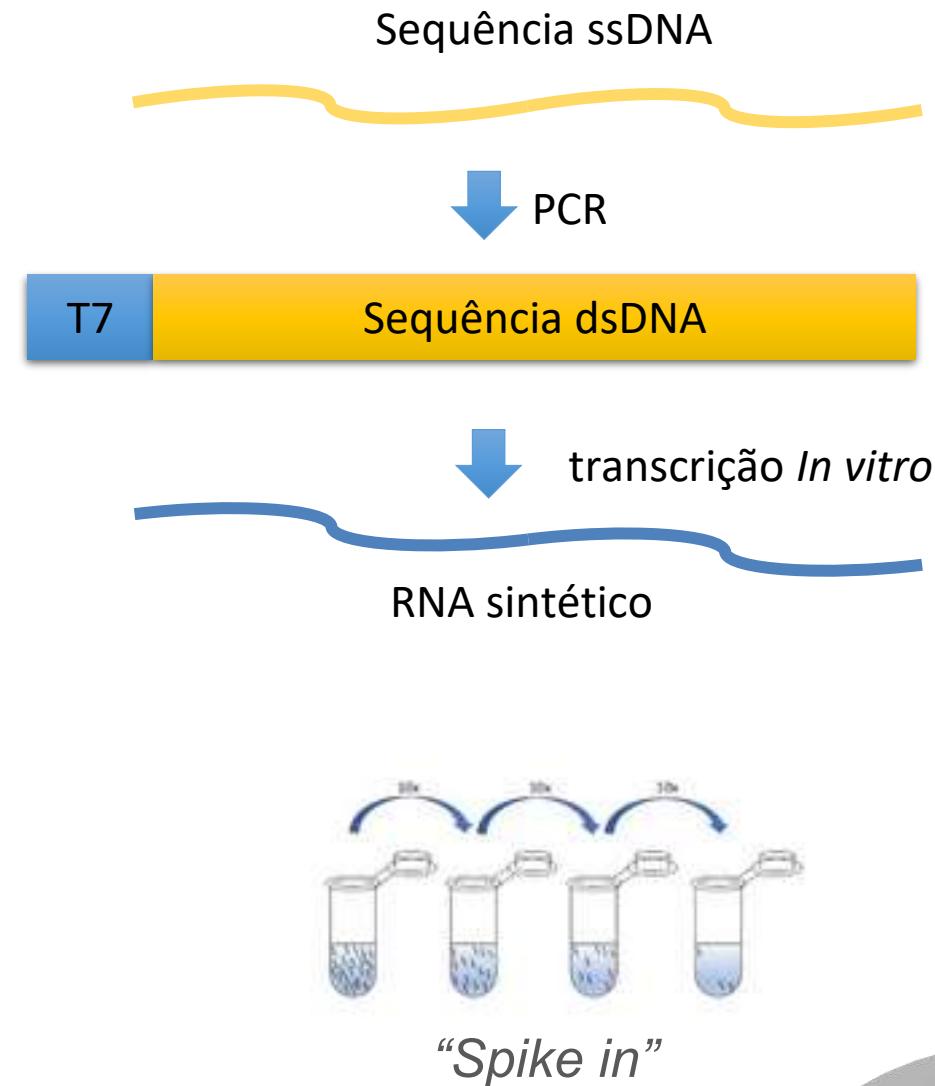
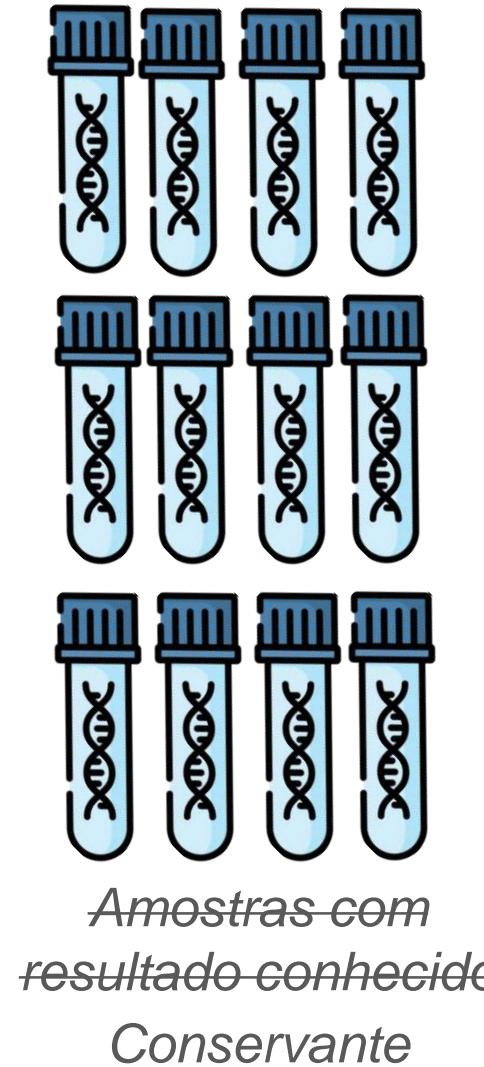


Diagnostics 2022, 12(7), 1539;

Pré-validação

# Material biológico e fase pré-analítica

N=60

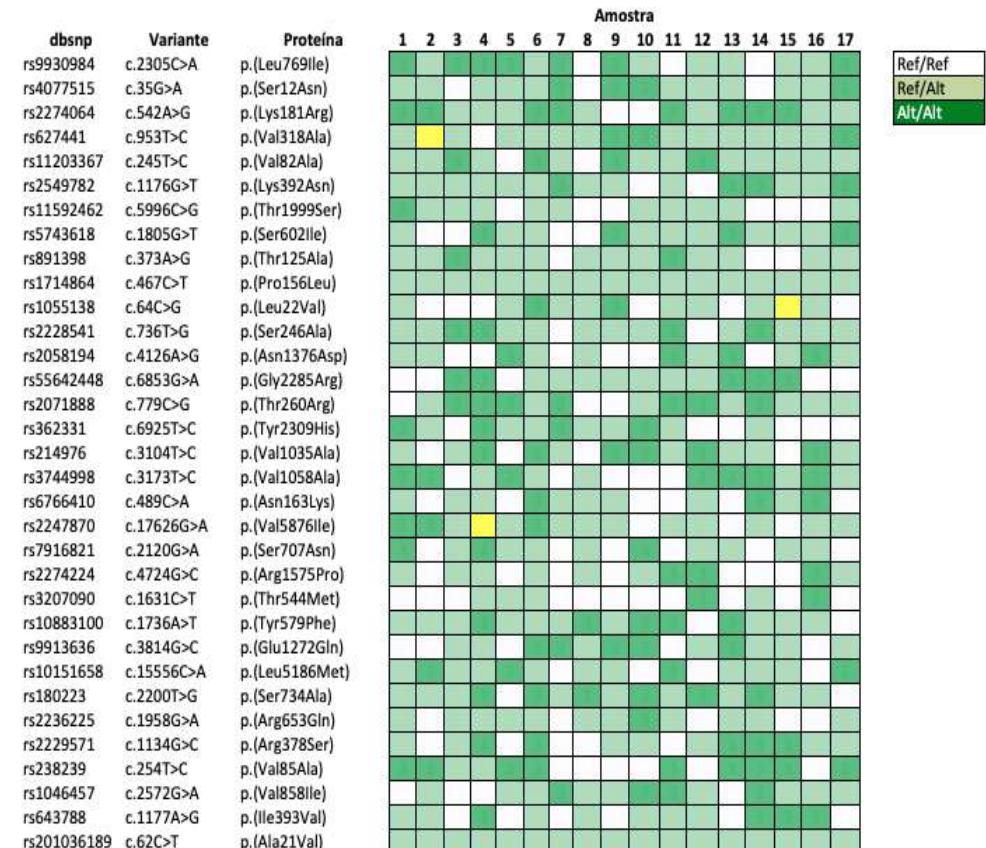
SARS-COV2  
2019

# Quantidade de amostras (n)

## qPCR

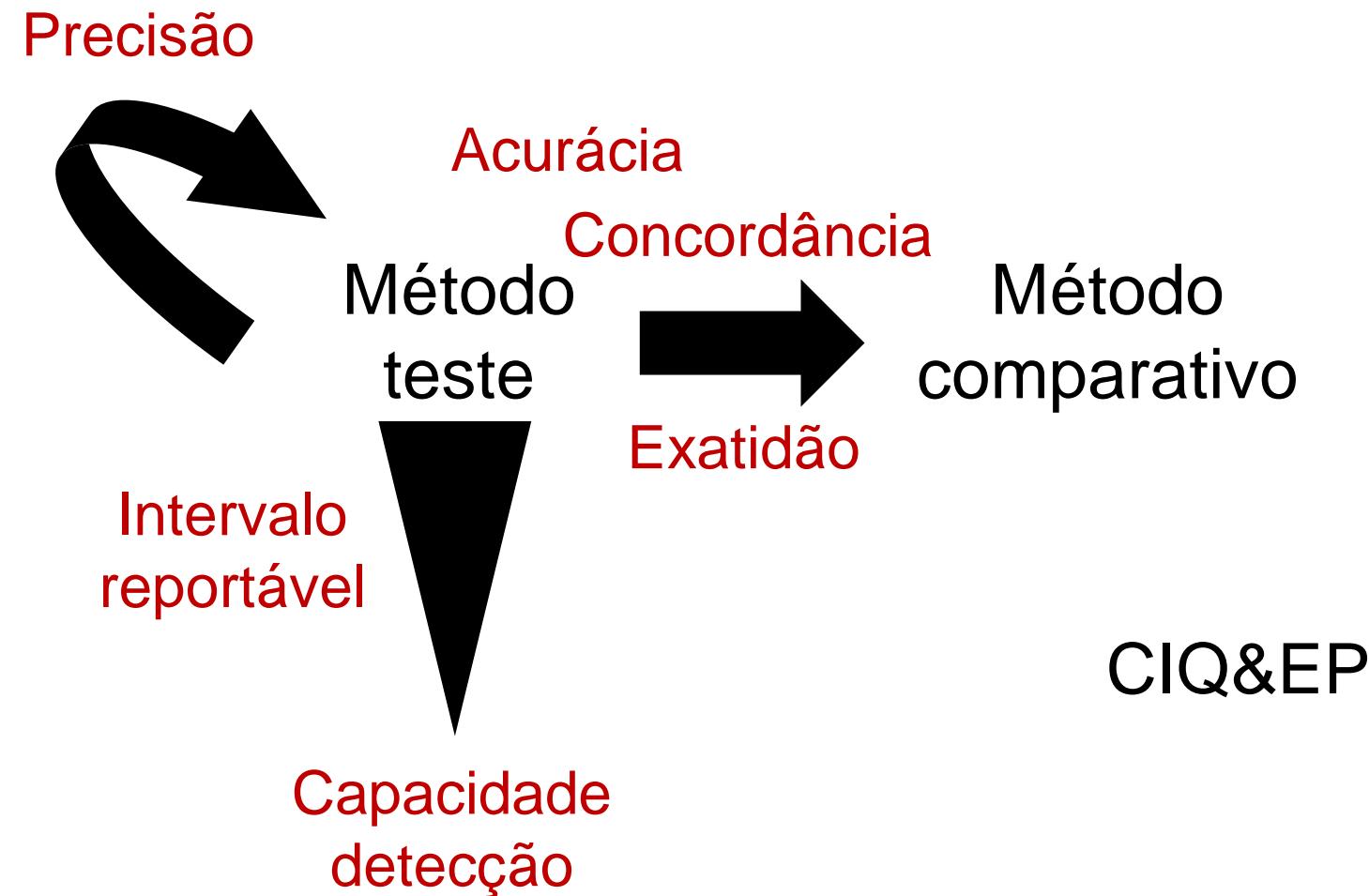
Tamanho amostral (n)	Concordância	Límite inferior 95% CI
10	100%	69,15%
20	100%	83,16%
30	100%	88,43%
40	100%	91,24%
50	100%	92,87%
60	100%	94,04%
300	100%	98,78%

## NGS



$n = 558/561 - 99,4\% - (95\% \text{CI } 98,4\text{-}99,9\%)$

# Parâmetros de desempenho



# Validação ensaio molecular qualitativo - exemplo

Open Access Article

## Analytical Sensitivity and Specificity of Two RT-qPCR Protocols for SARS-CoV-2 Detection Performed in an Automated Workflow

by  Gustavo Barcelos Barra <sup>\*,†</sup>  ,  Ticiane Henriques Santa Rita <sup>†</sup> ,  Pedro Góes Mesquita <sup>†</sup> ,  
 Rafael Henriques Jácomo  and  Lídia Freire Abdalla Nery 

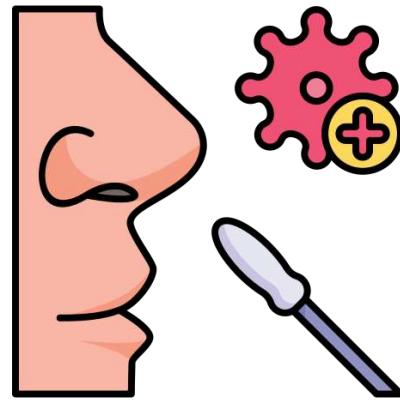
Research and development department, Sabin diagnostic medicine, Brasilia 70.632, Brazil

\* Author to whom correspondence should be addressed.

† These authors contributed equally to this work.

Genes **2020**, *11*(10), 1183; <https://doi.org/10.3390/genes11101183>

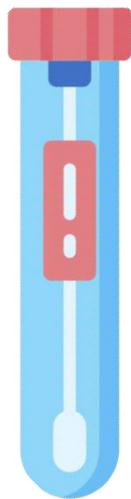
# Amostras



**Swab Nasal**

(n=60, saudáveis)

“Spike in”



RdRP

E  
N1  
N2

N and N3

T7 promoter

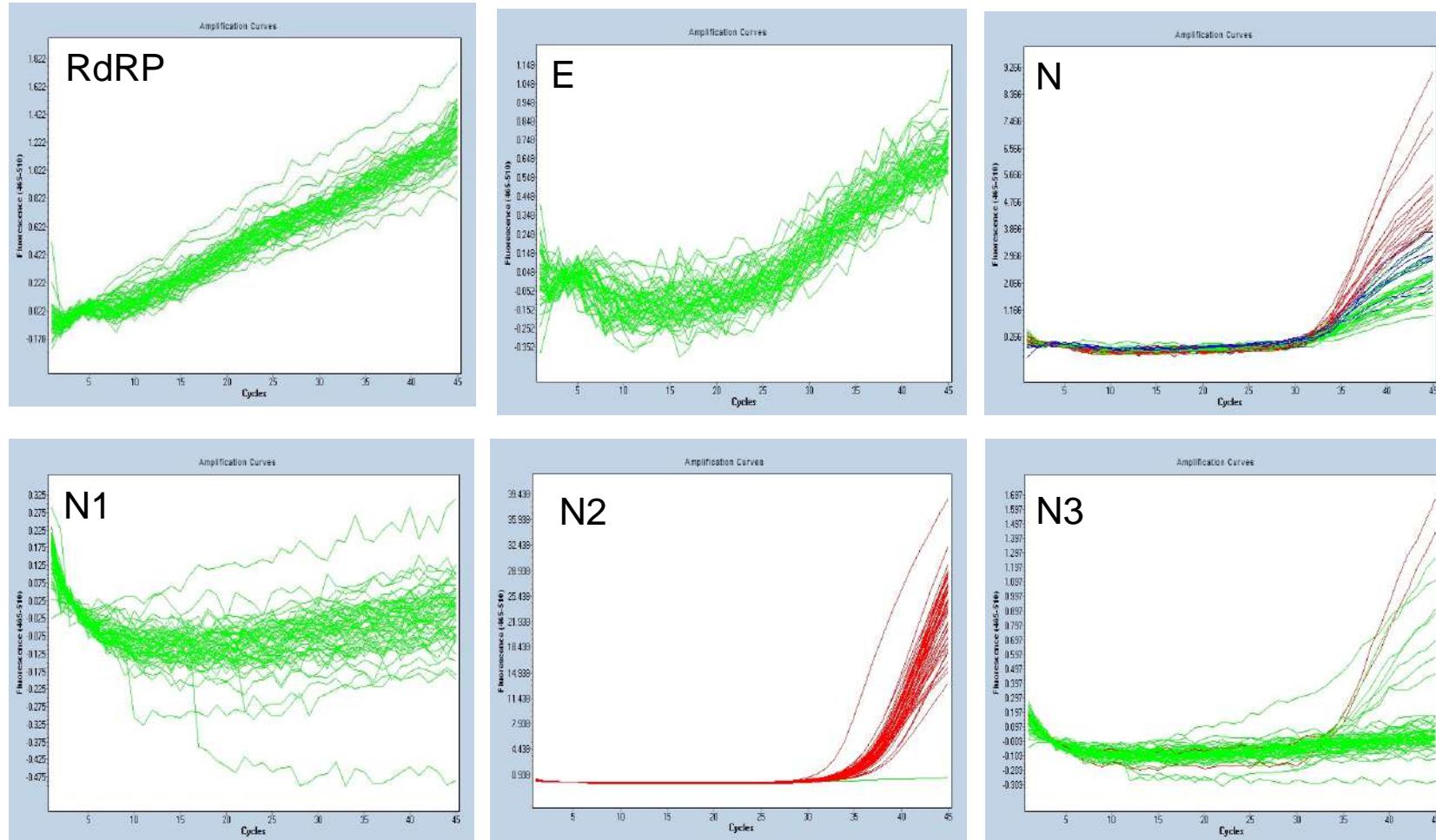
```

TAATACGACTCACTATAGGGGTGAAATGGTCATG
TGTGGCGGTTCACTATATGTTAACCAACCAGGTGGAA
CCTCATCAGGAGATGCCACAACGTGCTTATGCTAA
TAGTGTTTAACATTGACAGGTACGTTAATAG
TTAATAGCGTACTTCTTTCTTGCTTCTCGTGGT
ATTCTTGCTAGTTACACTAGCCATCCTTACTGCG
CTTCGATTGTGTGCGTACTGCTGCAATATGACCC
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GTGGACCCTCAGATTCAACTGGCAGTAACCAGAT
TACAAACATTGGCCGCAAATTGCACAAATTGCC
CCAGCGCTTCAGCGTTCTCGGAATGTCGCGCGG
GAGCCTTGAATACACCAAAAGATCACATTGGCAC
CCGCAATCCTGCTAACAAATGCTGCAATCGTGCTA
CAACTTCCTCAAGGAACAAACATTGCCAAAAGGCT
TCTACGCAGAAGGGAGCAGAGGCGGCAGTCAAGC
CTCTTCTCGTTCTC

```

Quantidade (cópias)

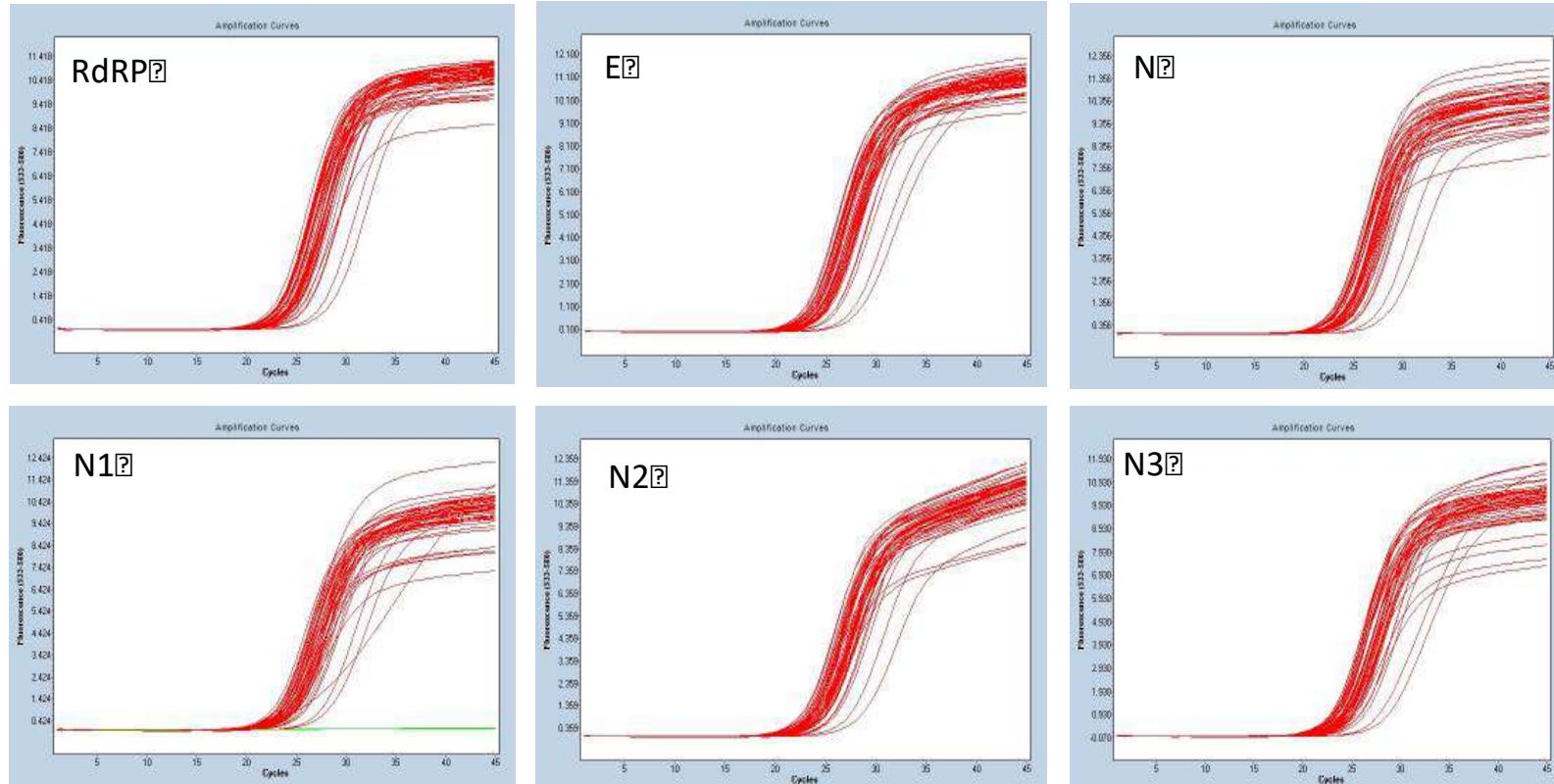
# Especificidade analítica - 6 ensaios



n=60, saudáveis

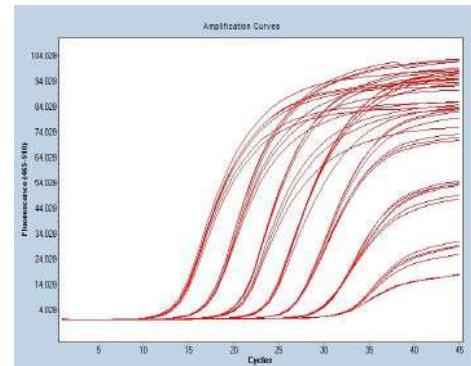
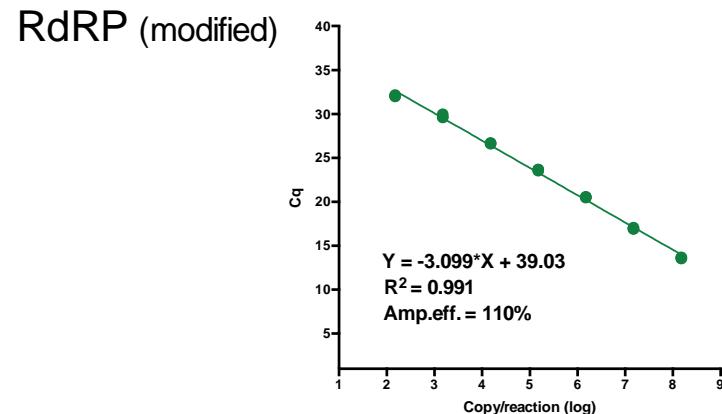
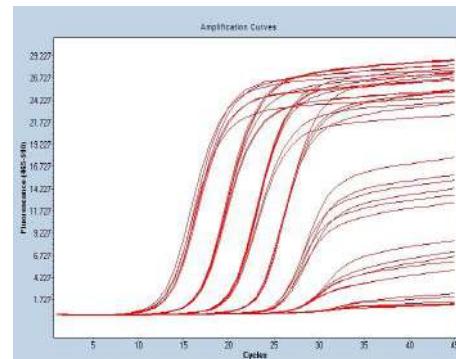
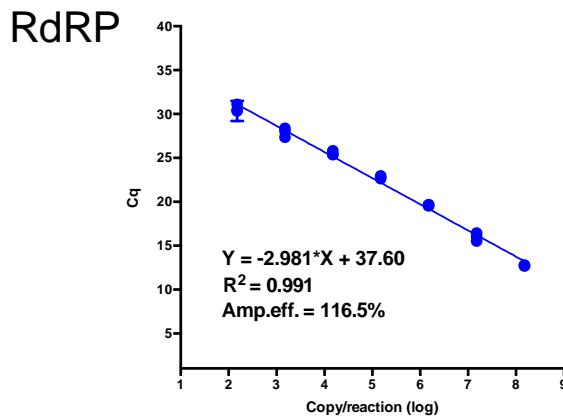
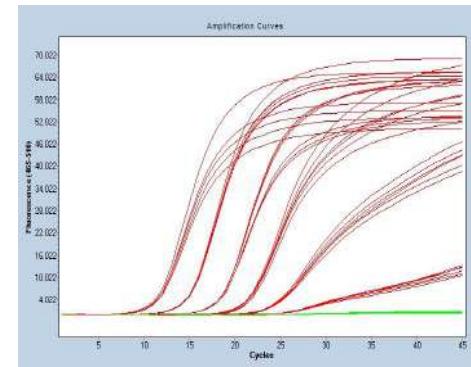
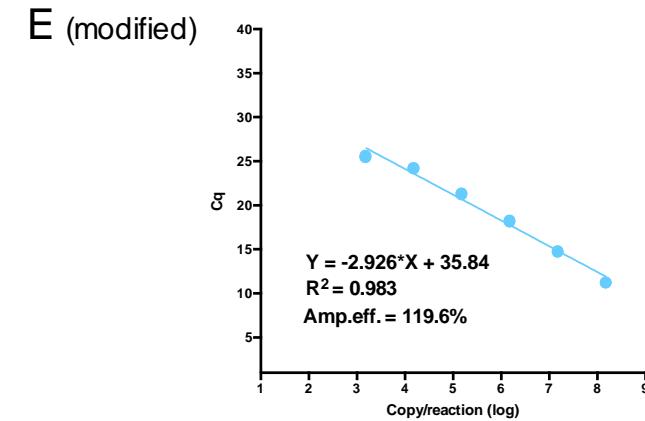
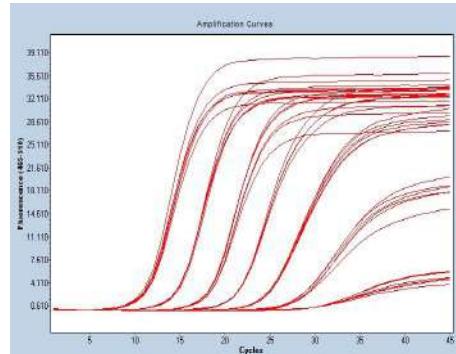
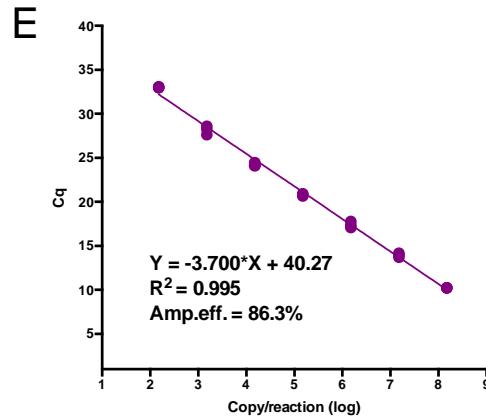
# Controle de reação – RNase P

Sample Control (RPP30) (HEX)



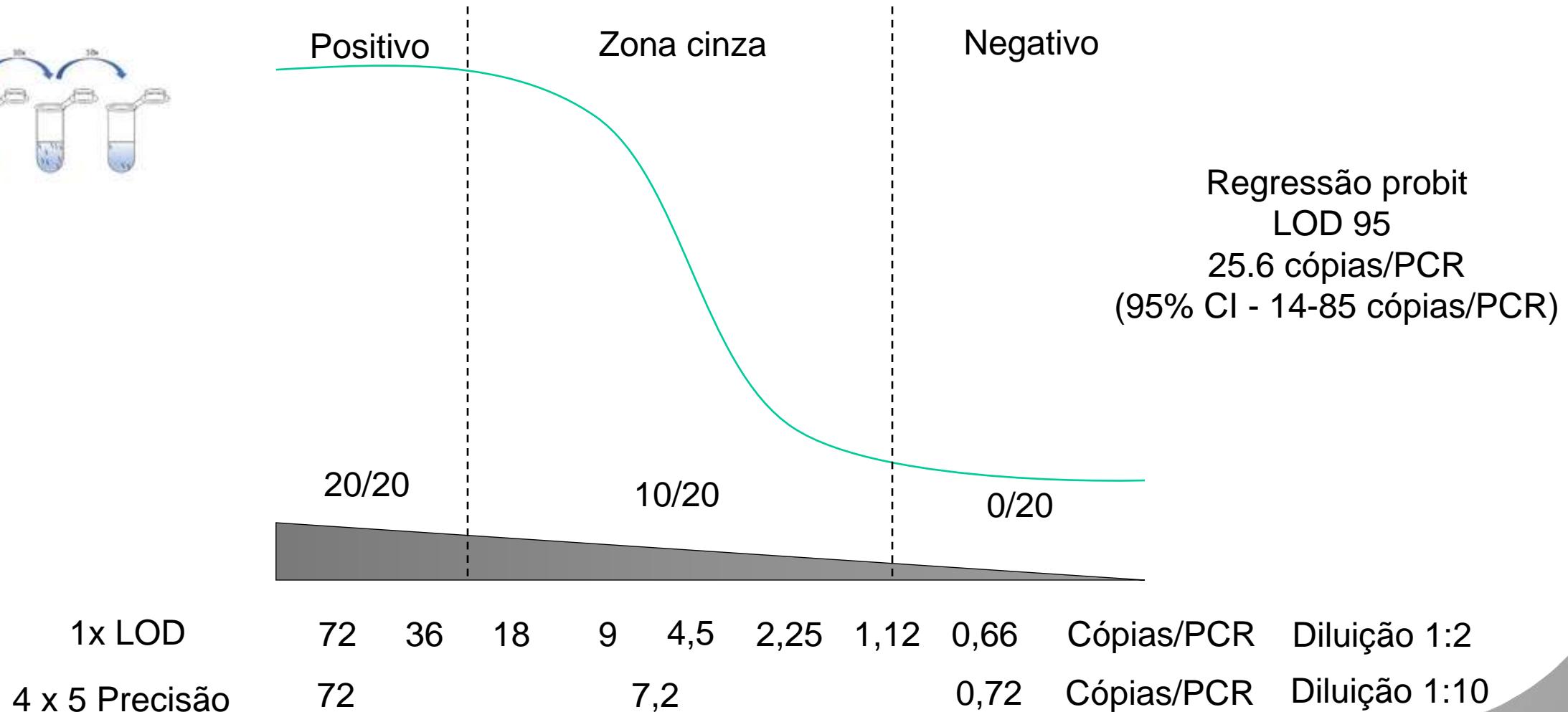
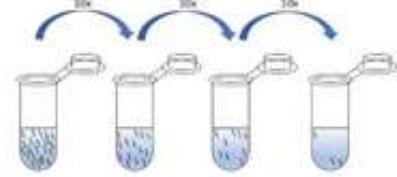
n=60, saudáveis

# Intervalo reportável – 1:10



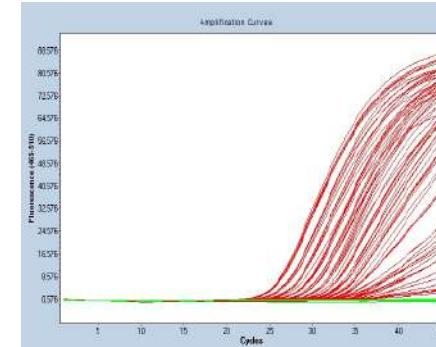
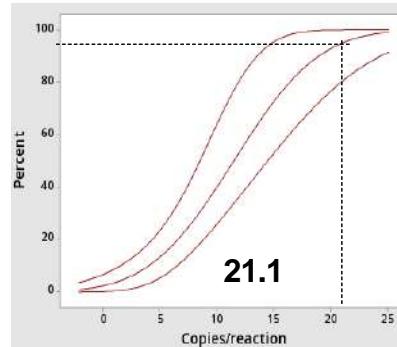
Alto, médio, baixo

# LOD e precisão - Qualitativo

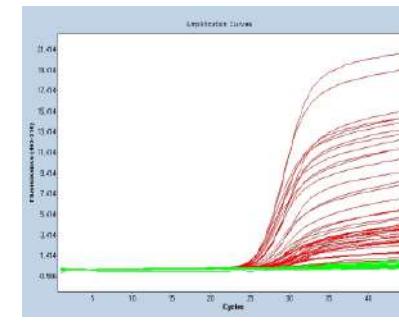
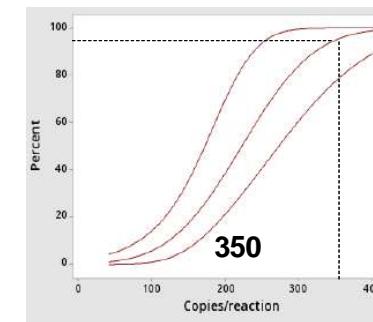


# LOD e precisão - Qualitativo

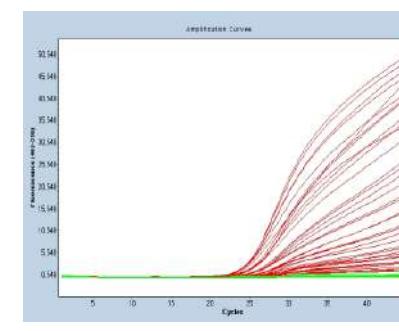
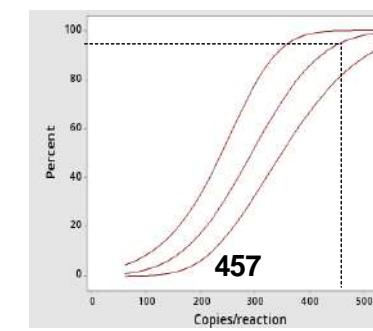
N1



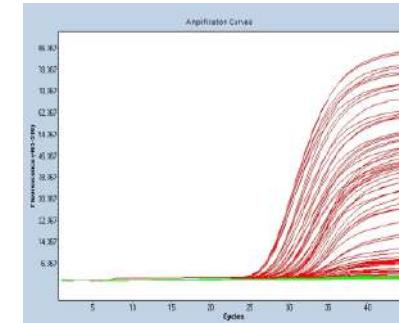
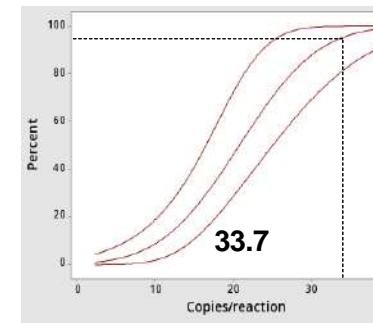
RdRP



E (modified)



RdRP (modified)



Regressão probit

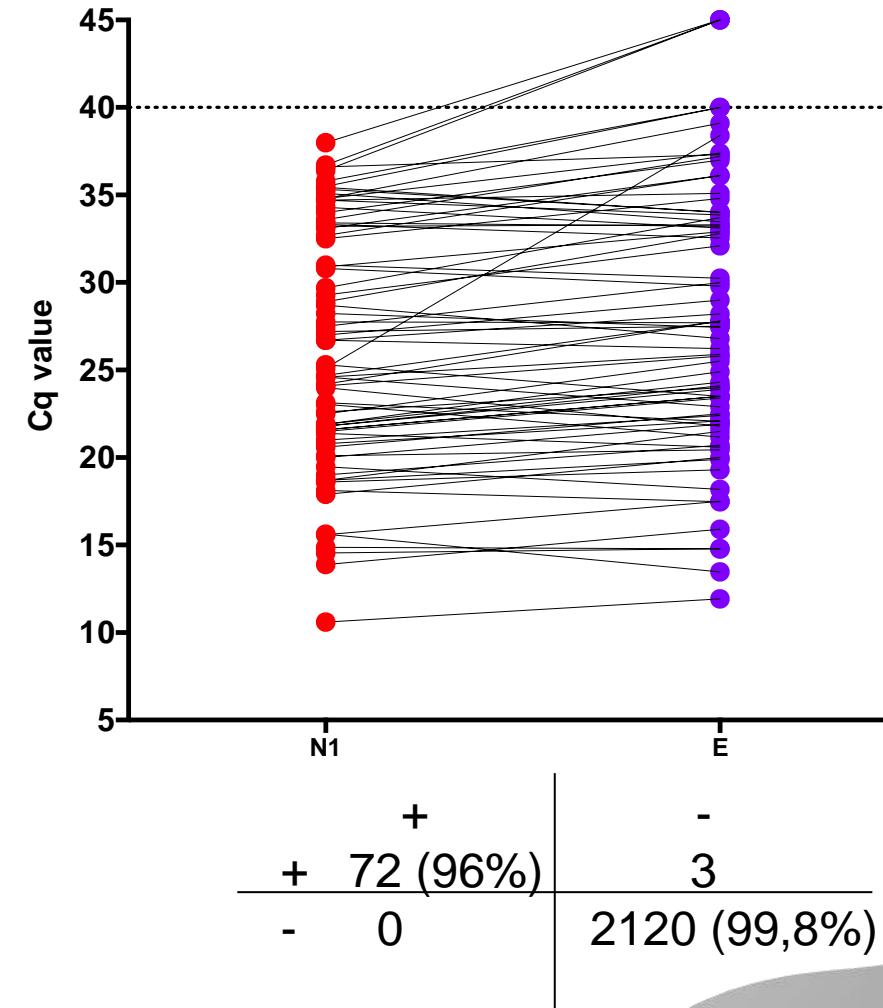
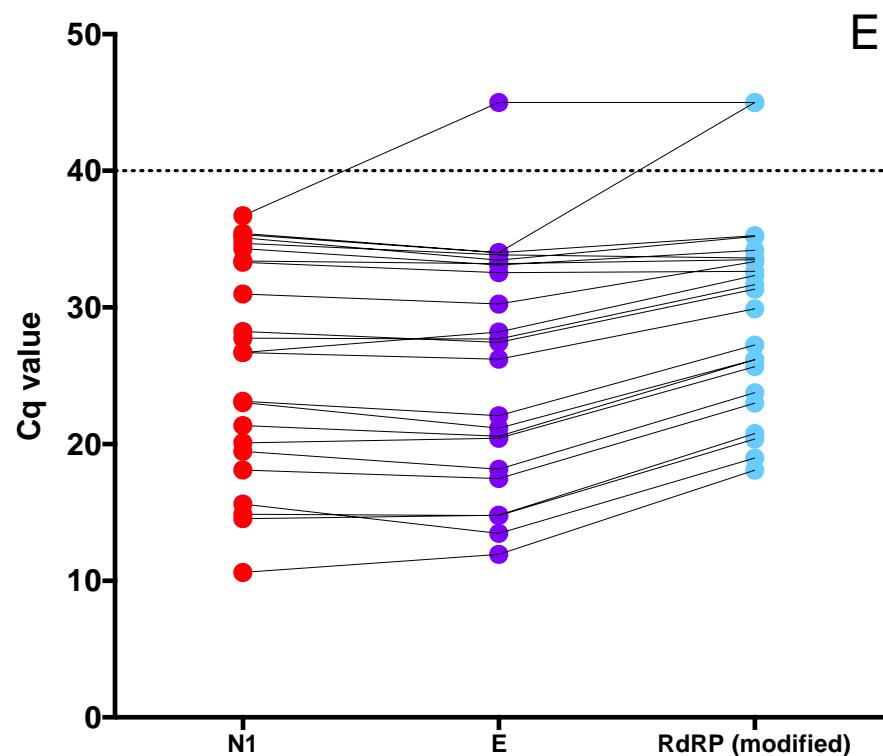
Diluição 1:2

# LOD e precisão - Qualitativo

SARS-CoV-2 diagnostic RNA Copies/Reaction	Tested replicates	Number of positive replicates				
		N1	E	E (modified)	RdRP	RdRP (modified)
26473,3	12	12	12	12	12	12
13236,6	12	12	12	12	12	12
6618,3	12	12	12	12	12	12
3309,2	12	12	12	12	12	12
1654,6	12	12	12	12	12	12
827,3	12	12	12	12	12	12
413,6	12	12	12	10	12	12
206,8	12	12	12	4	4	12
103,4	12	12	8	0	2	12
51,7	12	12	7	0	0	12
25,9	12	12	2	0	0	8
12,9	12	6	4	0	0	4
6,5	12	4	2	0	0	0
3,2	12	1	1	0	0	0
1,6	12	0	0	0	0	0
0,8	12	1	0	0	0	0
0,4	12	0	0	0	0	0
0,0	12	0	0	0	0	0

CIQ&EP

# Concordância on-going



# Overcoming Supply Shortage for SARS-CoV-2 Detection by RT-qPCR

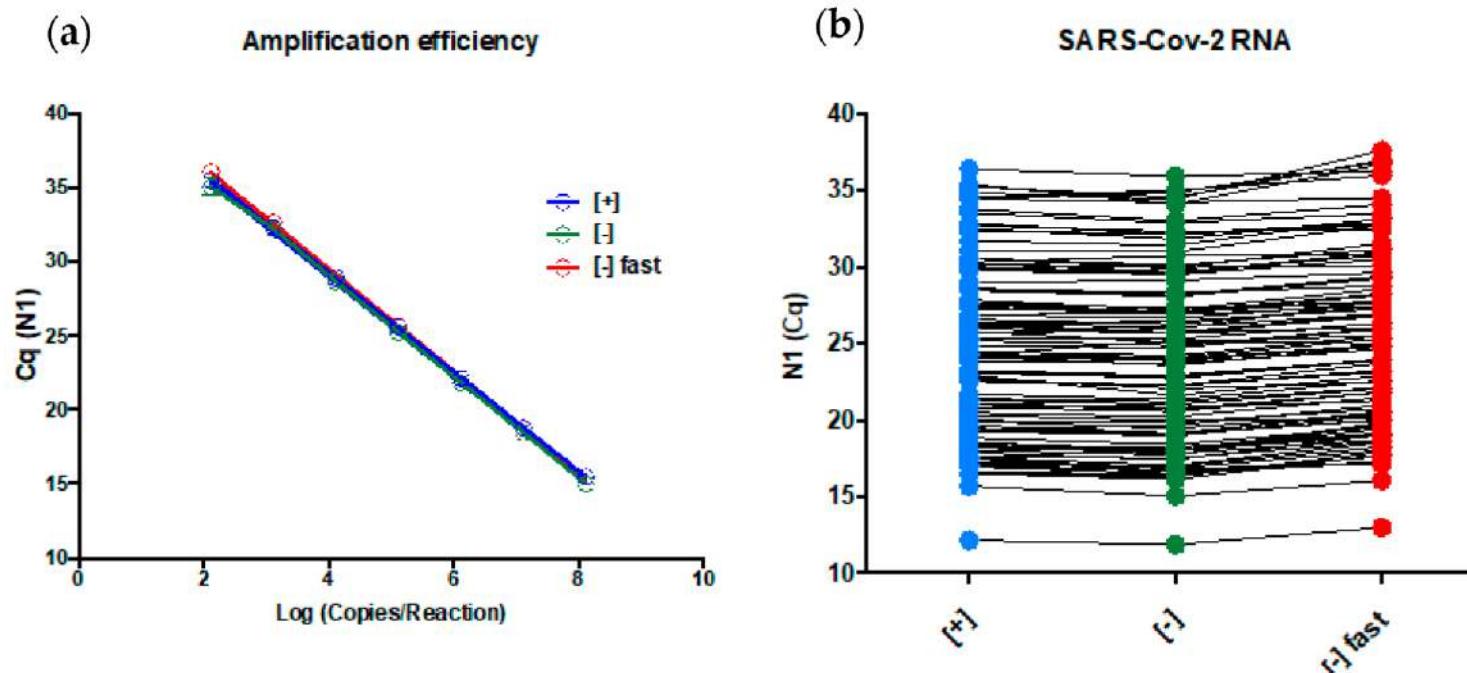
by  Gustavo Barcelos Barra \*<sup>†</sup> ,  Ticiane Henriques Santa Rita <sup>†</sup> ,  Pedro Góes Mesquita <sup>†</sup> ,  
 Rafael Henriques Jácomo  and  Lídia Freire Abdalla Nery 

Research and Development Section, Sabin Medicina Diagnóstica, 70632-340 Brasília, Brazil

\* Author to whom correspondence should be addressed.

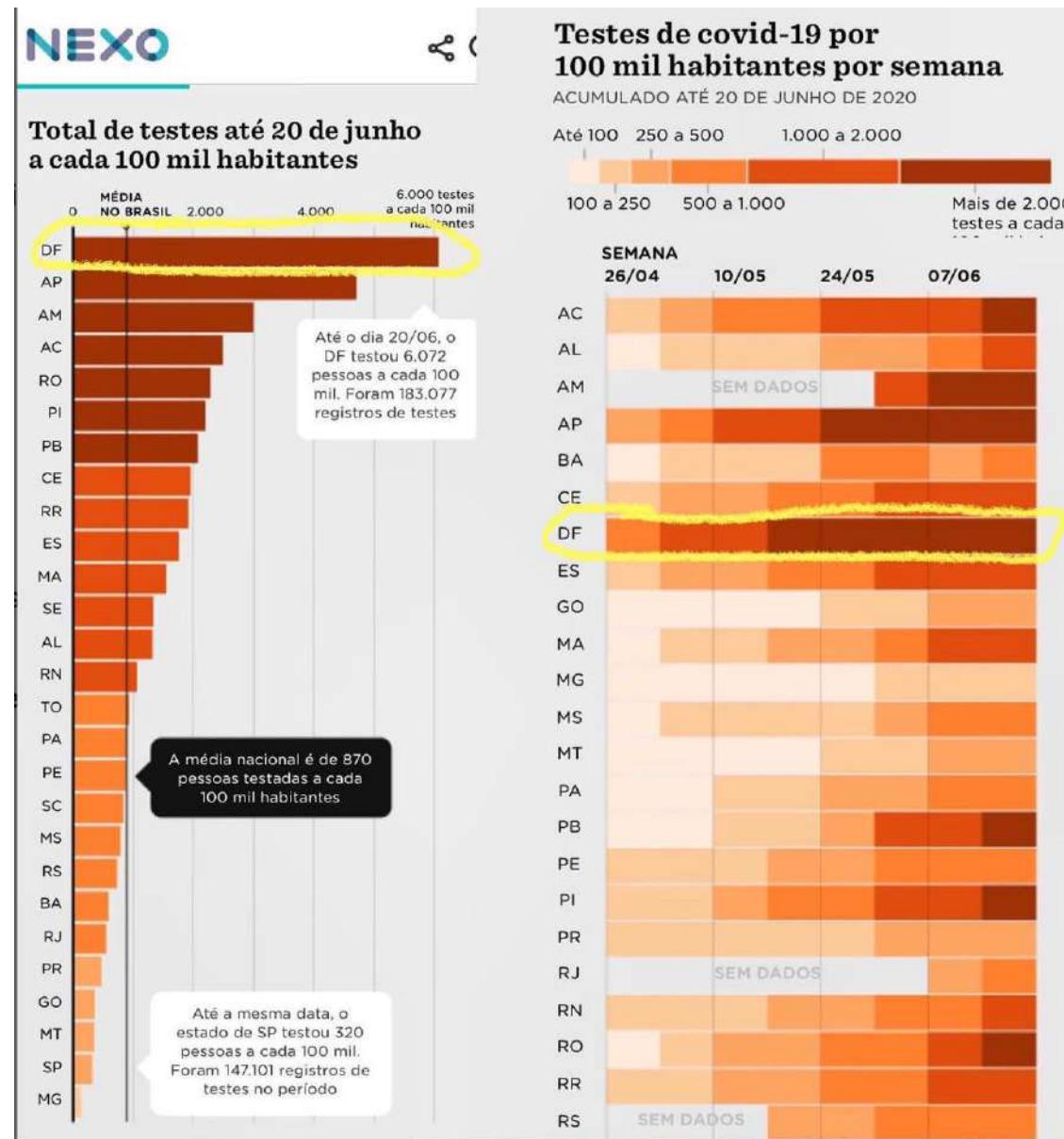
† These authors contributed equally to this work.

*Genes* **2021**, *12*(1), 90; <https://doi.org/10.3390/genes12010090>

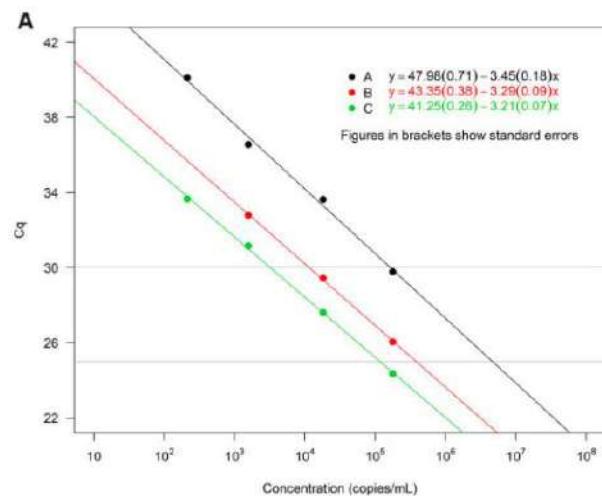
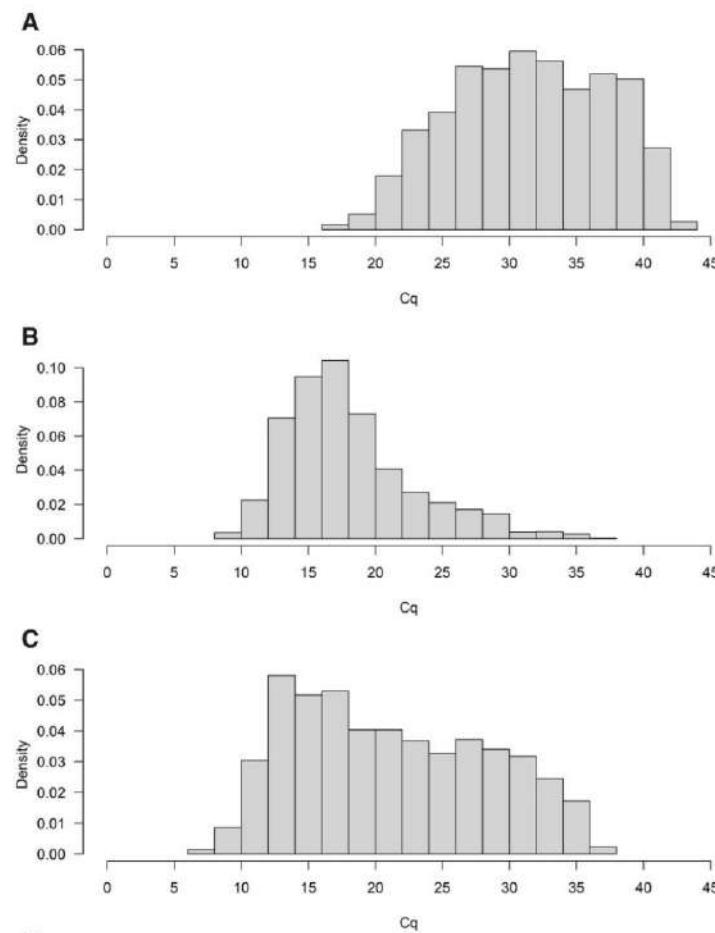


Produção > 30%  
Primer < 500%  
Probes < 250%

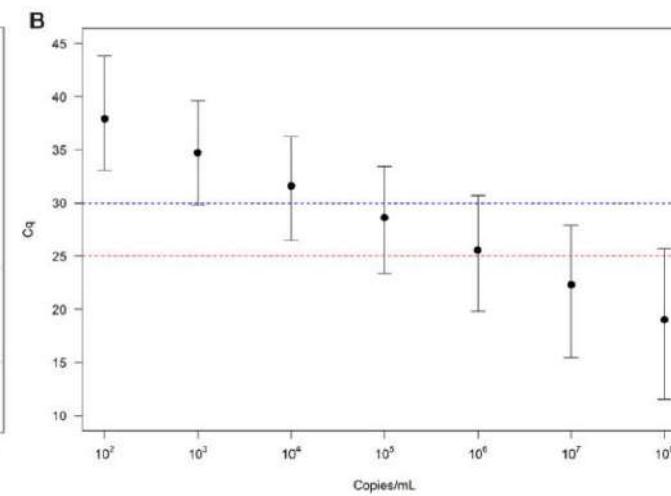
# Importância dos métodos próprios



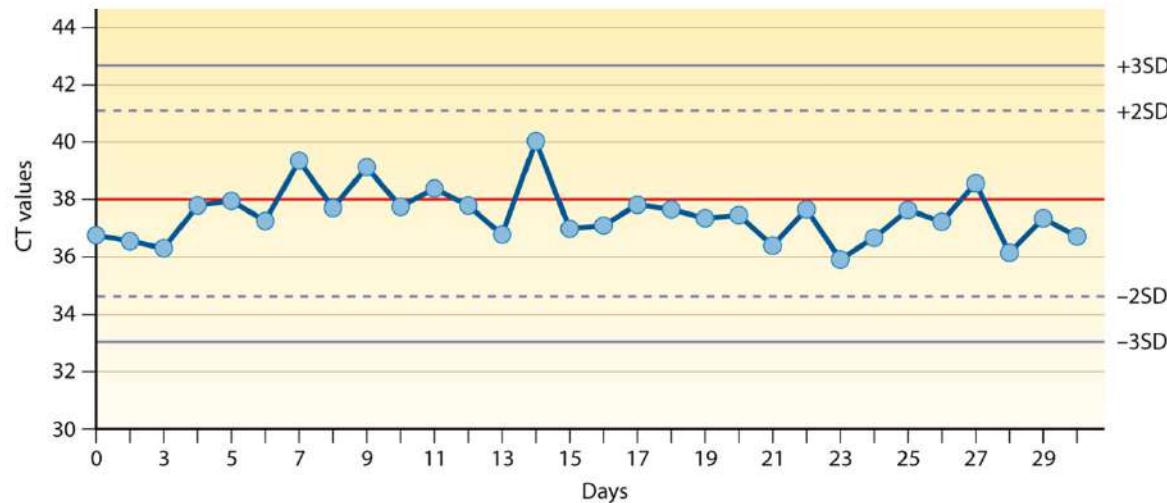
# Cq



Intervalo (99% CI) do valor  
Cq - EQA.



Intervalo (99% CI) do valor  
Cq - EQA.

**Valor**

Monitoramento contínuo do Cq do controle positivo baixo usando gráfico de Levey-Jennings plots.

**Desfecho**

#	Negativo	500 cópias/ml	Cq
1	Não detectado	Detetado	35,7
2	Não detectado	Detetado	36
3	Não detectado	Detetado	35,3
4	Não detectado	Detetado	37
5	Não detectado	Detetado	38
6	Não detectado	Não detectado	-
7	Detetado	Detetado	39/36

Monitoramento contínuo do desfecho do controle positivo baixo usando tabela simples

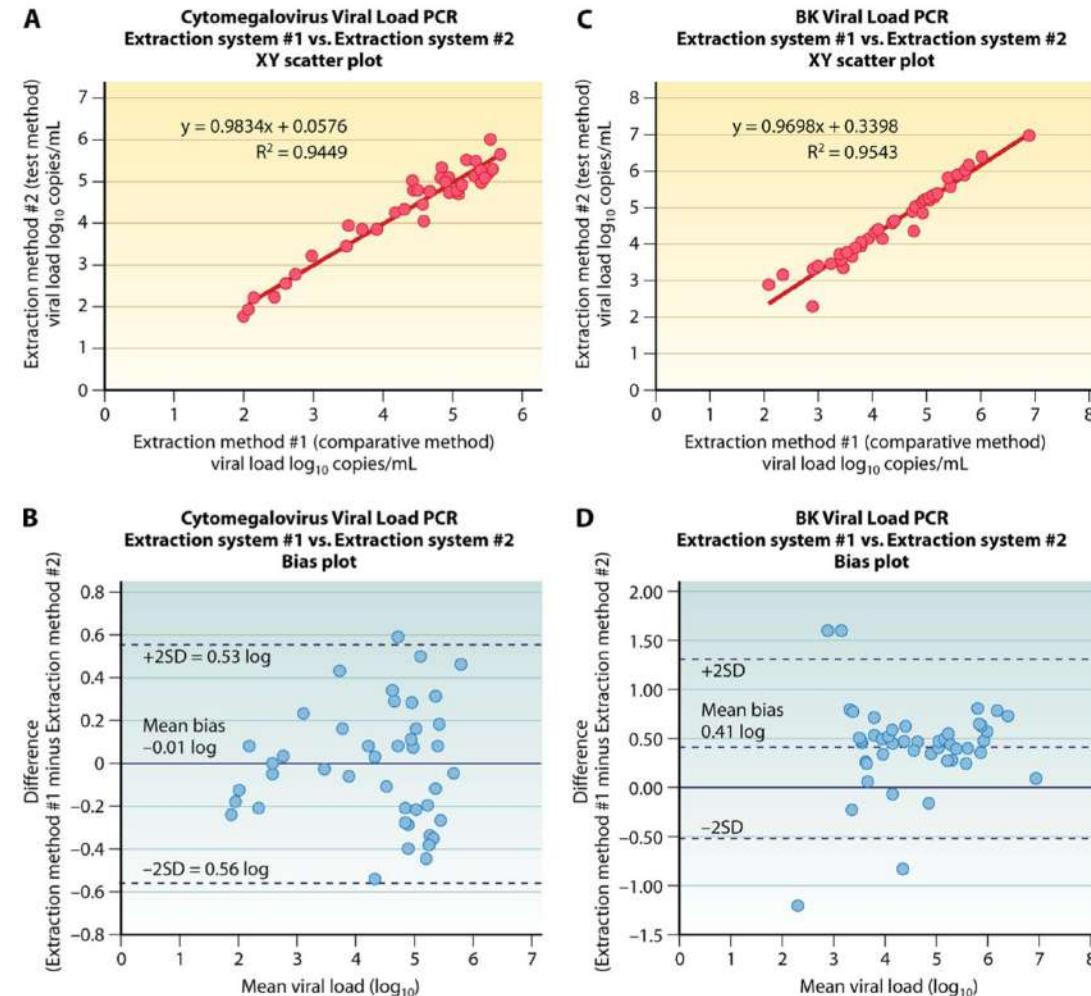
# Ensaios quantitativos

TABLE 2. Possible protocol for determining reportable range, analytical sensitivity, and precision in combined experiments based on current CLSI guidelines

Performance characteristic	Analyte concentration tested <sup>a</sup>											Comment(s)	
	Low					Medium			High				
	1	2	3	4	5	6	7	8	9	10	11		
Reportable range (for quantitative assays)	×	×	×	×	×	×	×	×	×	×	×	7-11 concentrations across anticipated measuring range; 2-4 replicates on same day	
Analytical sensitivity (LOD)	×	×	×	×	×							8-12 replicates of 4-5 samples at the low concentration end over 5 days	
Precision													
Qualitative assay	×	×	×									Use concentrations at LOD, 20% above LOD, and 20% below LOD; test in duplicate over 15 days (include data from analytical sensitivity runs to provide data over 20 days)	
Quantitative assay		×			×			×				Use high, low, and LOD concentrations; test in duplicate over 19 days (include data from reportable range study as day 1 to provide data over 20 days)	

<sup>a</sup> ×, the concentration is tested. The reportable range is from concentration 2 to concentration 10; the LOD, LLOQ, and upper limit of linearity are at concentrations 2, 4, and 10, respectively.

# Ensaios quantitativos - Exatidão



Burd EM. Validation of laboratory-developed molecular assays for infectious diseases. Clin Microbiol Rev. 2010 Jul;23(3):550-76. PMID: 20610823

# Ensaio de proficiência

LETTER TO EDITOR

J Bras Patol Med Lab, v. 53, n. 3, p. 194-195, June 2017

## Creation of a comprehensive proficiency testing program for molecular diagnosis in Brazil

*Criação de um amplo programa de ensaios de proficiência para o diagnóstico molecular no Brasil*

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10.5935/1676-2444.20170030

Gustavo B. Barra<sup>1</sup>; Rafael H. Jácomo<sup>1</sup>; Jéssica S. Gomes<sup>2</sup>; Rafael M. Lopes<sup>2</sup>

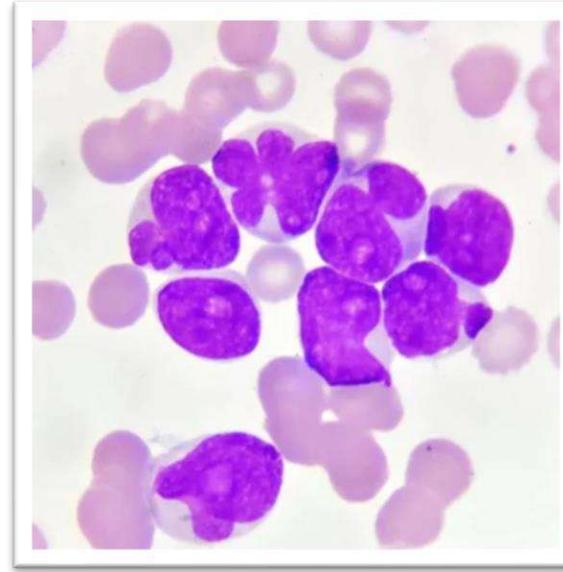
1. Laboratório Sabin de Análises Clínicas, Brasília, DF, Brazil. 2. Controllab, Rio de Janeiro, Brazil.

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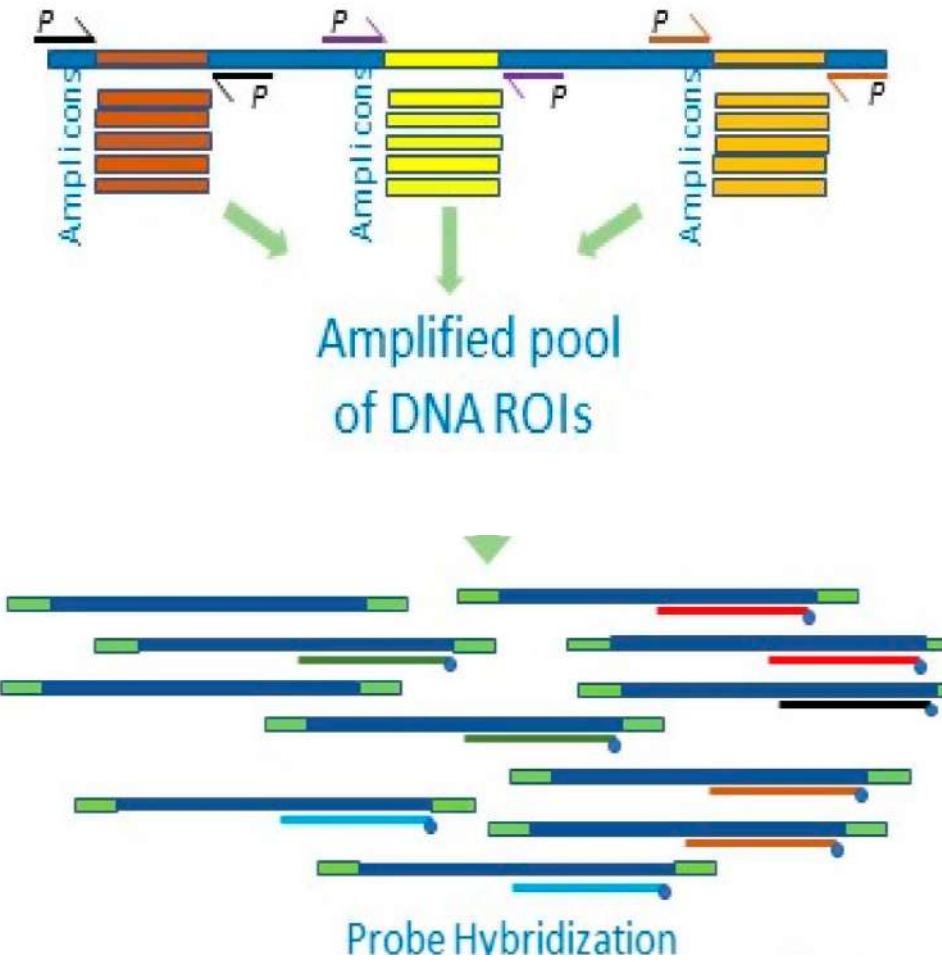
**Key words:** laboratory proficiency testing; molecular pathology.

# NGS – Painel mielóide

# Painel mieloide - amostras e ensaios



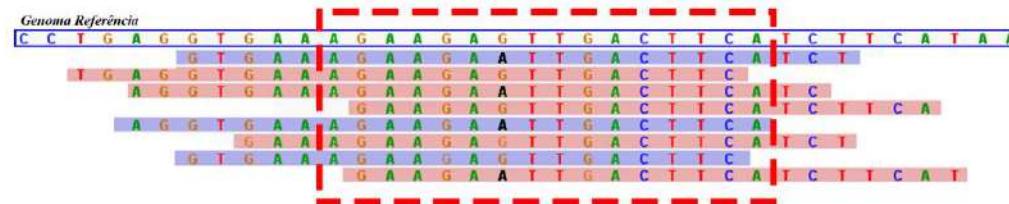
N=31, LMA



# Painel mieloide - intervalo reportável

40 genes

Gene	100X (%)	250X (%)	500X (%)		100X (%)	250X (%)	500X (%)
ABL1	100	100	100	KIT	100	100	100
ASXL1*	100	99,26	99,25	KRAS	100	100	100
BCOR*	97,46	97,46	95,93	MPL	100	100	100
BRAF	100	89,37	82,55	MYD88	100	100	100
CALR*	100	100	100	NF1*	100	100	100
CBL	100	100	100	NPM1	100	100	100
CEBPA*	100	100	100	NRAS	100	100	100
CSF3R	100	100	100	PHF6*	100	100	100
DNMT3A	100	100	100	PRPF8*	97,86	97,09	97,09
ETV6*	100	100	96,2	PTPN11	100	100	100
EZH2*	99,98	96,01	94,94	RB1*	96,52	96,52	96,5
FLT3	100	100	100	RUNX1*	100	100	100
FLT3ITD	100	100	100	SETBP1	100	100	100
FLT3TKD	100	100	100	SF3B1	100	100	100
GATA2	100	100	100	SH2B3*	100	96,85	91,36
HRAS	100	100	100	SRSF2	100	100	100
IDH1	100	100	100	STAG2*	97,33	97,33	93,66
IDH2	100	100	100	TET2*	100	100	100
IKZF1*	100	100	100	TP53*	100	99,68	99,36
JAK2	100	100	100	U2AF1	100	100	100
				WT1	100	100	100
				ZRSR2*	92,91	92,91	92,91





# Painel mieloide - Precisão

	Variante 1	T1	T2	Variante 2	T1	T2	Variante 3	T1	T2	Variante 4	T1	T2	Variante 5	T1	T2
1	-														
2															
3															
4															
5															
6															
7															
8															
9															
10															
11	DNMT3 Arg882His	0,47	0,46	IDH2 Arg140Gln	0,46	0,46	RUNX1 Ala324fs	0,43	0,43	CSF3R Ser783fs	0,06	0,06			
12	FLT3 Asp835Glu	0,07	0,07	NRAS Gly12Asp	0,14	0,15									
13															
14	WT1 Arg385fs	0,42	0,39	FLT3-ITD	0,19	0,19									
15	NPM1 Trp288fs	0,25	0,22	TET2 Gln758*	0,47	0,46	TET2 Gln769*	0,46	0,46	TET2 His922fs	0,42	0,44	ASXL1 Glu635fs	0,53	0,6
16	NPM1 Trp288fs	0,41	0,36	FLT3 Asp835Ala	0,08	0,09	NRAS Gly12Asp	0,16	0,16	FLT3 Ile836del	0,05	0,05	TET2 Tyr1560*	0,45	0,46
17															
18	NPM1 Trp288fs	0,39	0,44	WT1 Tyr407*	0,48	0,43	DNMT3A Leu508fs	0,47	0,4	FLT3 Val592Asp	0,41	0,4			
19															
20															
21	NPM1 Trp288fs	0,41	0,43	DNMT3A Arg882His	0,45	0,46									
22															
23															
24															
25															
26															
27															
28															
29															
30	-														

n=9, 7 positivas (24 variantes) e 2 negativas (sem variantes)

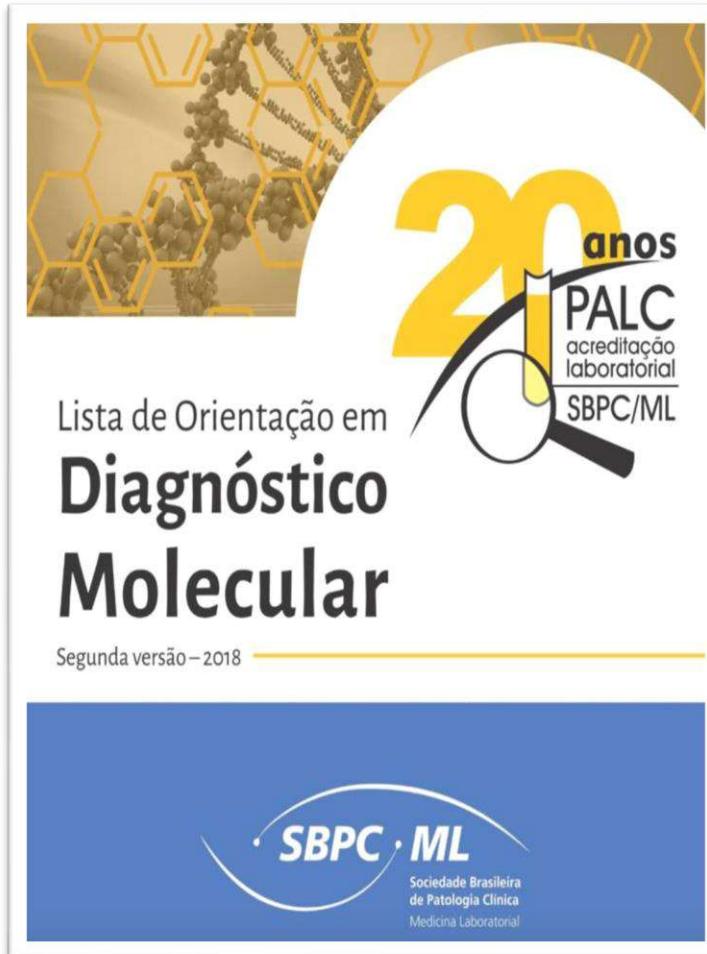
**Amostras 9**  
Positivas 7  
Negativas 2

**Variantes 24**  
24/24 T1 & T2

**On-going**



# Lista de orientação em diagnóstico molecular SBPC/ML





EP

Obrigado e  
até a próxima!



**Gustavo Barra**

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